THE SPACE STRUCTURES OF α-MELANOTROPIN

G. V. NIKIFOROVICH, M. D. SHENDEROVICH and G. I. CHIPENS

Institute of Organic Synthesis, Latvian SSR Academy of Sciences, 21 Aizkraukles, Riga 226006, USSR

Received 9 February 1981

1. Introduction

The study of conformation—function relationships in biologically active peptides requires a detailed knowledge of their space structure. Semi-empirical conformational analysis is one of the methods providing the necessary information [1–4]. This communication describes the set of low-energy structures of α -melanotropin (α -MSH) peptide backbone determined by that method. α -MSH (Ac-Ser₁—Tyr₂—Ser₃—Met₄—Glu₅—His₆—Phe₇—Arg₈—Trp₉—Gly₁₀—Lys₁₁—Pro₁₂—Val₁₃—NH₂) is the natural oligopeptide which in many respects can be regarded as a natural analogue of corticotropin (ACTH).

2. Methods and results

The intramolecular conformational energy U involved the non-bonded, electrostatic and torsional potentials along with the hydrogen bond potentials [4]. The ionogenic side chains of the molecule (Glu, Arg, Lys) are assumed completely ionized [5]. The sets of local energy minima of the peptide backbone, i.e., B ($\varphi \sim -140^\circ$, $\psi \sim 140^\circ$), R ($\varphi -60^\circ$, $\psi \sim -60^\circ$), L ($\varphi \sim 60^\circ$, $\psi \sim 60^\circ$) and H (for the Gly₁₀ residue; $\varphi \sim 80^\circ$, $\psi -80^\circ$) conformations and all the $X_1 \sim 60^\circ$, 180° , -60° angle rotamers were considered as possible molecule conformations. The values of dihedral angles $X_2 - X_4$ were chosen in accordance with the calculation results obtained for the appropriate monopeptides [6]. The principal steps involved in the selection of low-energy backbone structures are as follows:

(1) Determination of the set of low-energy backbone structures for molecule fragment 1-6 using the conformational energy U calculations for all possible di- and tripeptide conformations and consecutive estimation of 'near-neighbouring' interaction energies in tetra-,

- penta- and hexapeptide fragments (see [2]). The final stage involves the refinement of side chain spacing for each of the selected hexapeptide backbone structures using an algorithm for the selection of energetically optimal dihedral angles X [7]. As a result, 46 backbone structures of fragment 1-6 were found to meet the requirement $\Delta U = U U_{\min} \le 10 \text{ kcal/mol}$.
- (2) The preliminary evaluation of the ionized group electrostatic interaction energy, Eel, and the energy of 'near-neighbouring' interactions in the backbone, Eb, resulted in ~750 structures of the fragment 5-11 backbone for which $\Delta(E^{b} + E^{el}) \leq 20 \text{ kcal/mol.}$ At the same time, only 84 of them appear to satisfy the criterion $\Delta U \leq 15$ kcal/mol, as judged by the results of energy U calculations for the structures of the 'model fragment': Glu-Ala-Ala-Arg-Ala-Gly-Lys. The optimal backbone conformations of fragment 12-13 have been determined for each of the 84 structures based on the calculation results for the fragment 7–13 (selection of the backbone structures satisfying the criterion $\Delta U \leq 10$ kcal/mol according to the scheme: $8-11 \rightarrow 8-13 \rightarrow 7-13$ accompanied by the refinement of the side chain spacing at each step). This was followed by the determination of 34 backbone structures for the fragment 5–13 which meet the requirement $\Delta U \le 15$ kcal/mol when the side chains are optimally spaced.
- (3) All the possible variants of the fragment 1-4 backbone contained in the earlier selected set of fragment 1-6 backbone structures were examined for each of 34 backbone structures of fragment 5-13 at the level of complete α -MSH molecule. The final step of the calculations including the refinement of side chain spacing led to the selection of 36 (out of > 200 calculated) types of low-energy backbone structures of α -MSH ($\Delta U \le 12$ kcal/mol). Table 1 demonstrates

structures characterized by the optimal backbone conformation of fragment 1 4 with respect to backbone structure of fragment 5-13. Structure 1 from table 1 is depicted in fig.1.

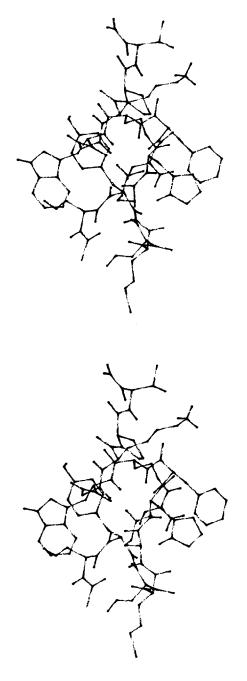


Fig.1. The α -melanotropin structure with lowest energy.

3. Discussion

The most remarkable feature of the structures presented in table 1 is the close spacing of the side chains of Glu₅ and Arg₈ residues. At the same time,

Table 1 The set of low-energy α -MSH structures

Backbone structures Residue Angle 1 -128 -126 -133 -133 -111 -119 -123 126 120 142 118 142 136 111 176 176 -179 179 180 175 179 176 162 180 -121 -132 -55 -53 31 -53 11 142 166 -162 - 57 -78 -61 -80 96 97 101 -88 -83 -151 -107 -97 -144 -107 -114 -128 -115 -53 - 34 -35 -50 -33 - 37 142 -32 - 30 -48 -53 58 52 180 180 179 178 180 178 179 -175 -102 -127 -124 -113 -121 -105 -137 -105 -127 158 153 110 132 149 -160 -64 -160 -81 -61 -101 +178 -161 178 180 168 178 -178 180 175 -177 -178 -179 171 -119 -105 -109 -135 -152 -114 -133 120 132 144 131 122 127 152 150 133 Glu -126 -122 -168 5.7 169 -66 -67 -101 -177 86 86 -131 -148 -150 -136 -135 -102 -104 110 -28 - 39 -41 40 -44 - 58 40 -42 -61 81 -80 -81 -80 -80 -81 . 80 -137 -122 -138 -53 31 -116 -110 -141 -37 -41 136 -58 -76 -76 -74 -61 -42 -40 -57 179 100 92 101 104 91 -92 -103 -135 -117 -112 -118 -140 -105 -60 -70 -47 -53 -71 -70 -78 -66 -73 -73 -78 -67 -73 -76 -78 145 -80 -81 171 -81 -154 -139 -151 -78 -160 173 -159 111 143 126 144 104 109 -84 105 145 140 89 -114 -81 -154 -128 -153 -117 127 152 129 144 143 Trp 180 178 178 -76 -81 77 8.3 85 -100 68 83 -101 -78 98 101 Gly 51 - 38 -109 -130 -105 -124 -135 -146 -123 -111 -125 -132 -131 -86 -143 137 121 133 135 102 130 113 136 143 111 146 140 142 -171 179 175 -171 162 162 147 163 180 170 178 156 -169 -172 166 165 172 175 164 180 171 -31 - 39 110 140 129 -124 -134 -106 -137 -123 -110 -106 -120 -178 176 180 -179 -179 -177 -178178 -176 -177 Interatomic Colu-Carg 3.8 3.4 3.1 3.3 distances, Cd Lys 8.8 3.5 3.0 9,9 10.7 Structure of

fragment 6-9

the side chain of lysine is in many cases directed 'outside' the fragment 6-9 (e.g., fig.1) which can acts as the 'active centre' of the molecule and allegedly provides direct binding to specific receptors [8]. This peculiarity of the molecule's space organization agrees well with the high melanotropic activity of α -MSH analogues, where Lys₁₁ is substituted by Nle, Ser or even Gly [9,10]. Furthermore, it can elucidate the reasons for the drop in lipolytic activity found for ACTH 2-19 analogues with cystine bridges of (2,10), (3,10) or (5,10) type [11]. Typically, the data in table 1 frequently imply the retention of BRRB or BRRR structures for the fragment 6-9 backbone, thus indicating considerable conformational rigidity of this 'active centre'.

The data given in table 1 are also in keeping with the results of physico—chemical studies on the ACTH space structure which indicate the presence of α -helix elements [12] in the peptide backbone as well as with the estimation of experimental values (\sim 10 Å) found for the distance between the side chains of Tyr₂ and Trp₉ residues [13]. Consequently, it can be assumed that the set of low-energy conformations found in this study contains sufficient information concerning the main features of α -MSH space organization and can be therefore applied to the study of conformation—function relationships for α -melanotropin and adreno-corticotropin.

References

- [1] Scheraga, H. A. (1968) Adv. Phys. Org. Chem. 6, 103-183.
- [2] Galaktionov, S. G., Nikiforovich, G. V., Shenderovich, M. D., Chipens, G. I. and Vegner, R. E. (1976) in: Peptides-1976 (Loffet, A. ed) pp. 617-624, Bruxelles.
- [3] Balodis, Yu. Yu., Nikiforovich, G. V., Grinsteine, I. V., Vegner, R. E. and Chipens, G. I. (1978) FEBS Lett. 86, 239-242.
- [4] Nikiforovich, G. V., Leonova, V. I., Galaktionov, S. G. and Chipens, G. I. (1979) Int. J. Pept. Prot. Res. 13, 363-373.
- [5] Nikiforovich, G. V., Rosenblit, S. A. and Chipens, G. I. (1980) in: abst. 3rd USSR-FRG Symp. Peptide Chemistry, p. 39, Moscow.
- [6] Akhrem, A. A., Golubovich, V. P., Galaktionov, S. G., Nikiforovich, G. V., Shenderovich, M. D. and Sherman, S. A. (1976) Izv. Akad. Nauk BSSR, chem. ser. 5, 82-93.
- [7] Nikiforovich, G. V., Shenderovich, M. D. and Balodis, Yu. Yu. (1981) Bioorg. Khim. in press.
- [8] Otsuka, H. and Inouye, K. (1964) Bull. Chem. Soc. Japan 37, 1465--1471.
- [9] Eberle, A. N. (1976) Dissertation ETH 5735, Zurich.
- [10] Geiger, R., Sandow, J. and Kastin, A. J. (1977) Hoppe-Seyler's Z. Physiol. Chem. 358, 1475–1481.
- [11] Blake, J., Rao, A. J. and Li, C. H. (1979) Int. J. Pept. Prot. Res. 13, 346-352.
- [12] Low, M., Kisfaludy, L. and Fermandjian, S. (1975) Acta Biochim. Biophys. Acad. Sci. Hung. 10, 229-231.
- [13] Eisinger, J. (1969) Biochemistry 8, 3902-3908.
- [14] IUPAC-IUB Commission on biochemical nomenclature (1974) Pure Appl. Chem. 40, 293-307.